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DESIGN AND SYNTHESIS OF HETEROCYCLIC ANALOGUES OF MYCOPHENOLIC ACID AS POTENTIAL CHEMOTHERAPEUTIC AGENTS

Jeewoo Lee1* and Wayne K. Anderson*

Department of Medicinal Chemistry, School of Pharmacy, State University of New York at Buffalo, Buffalo, New York, 14260

Abstract: A series of heterocyclic analogues **3-10** of mycophenolic acid were designed as metabolically stable congeners possibly targeted for inosine monophosphate dehydrogenase. The synthesis, cytotoxic and anti-HIV activity of these compounds are described.

Mycophenolic acid (MPA) 1, NSC 129185, is a potent inhibitor of inosine monophosphate dehydrogenase (IMPD, IMP:NAD⁺ oxidoreductase, EC 1.1.1.205), a rate-limiting enzyme in guanine nucleotide biosynthesis.² The compound may bind in the NAD⁺/NADH domain of the IMPD active site similar to other IMPD inhibitors such as tiazofurin 2, NSC 286193, and selenazofurin.³ It has been shown to possess antineoplastic, antifungal, antiviral, antiinflammatory, and immunosuppressive activity.⁴ The synthesis and structure activity relationship of MPA have been extensively studied.^{4,5,6} Attractive features of this drug include low toxicity and the reversibility of toxicity upon withdrawl of drug. However, a major problem with MPA in humans is the rapid conjugation of the C-7 phenolic hydroxyl group with glucuronic acid, thereby preventing the accumulation of therapeutic levels of the drug.⁷

As one approach toward solving the problem of conjugation, we tried to replace the metabolically labile phenolic hydroxyl in MPA with more stable moieties which can act as hydrogen bonding groups. First, the phenolic hydroxyl of MPA was replaced by bio-isofunctional heterocyclic groups such as pyrrole 3-4 and pyridine 5 which can function as either a hydrogen bond acceptors or donors in the active site of enzyme.

These heterocycles are called "fixed geometry" hydrogen bonding groups because the colinear hydrogen geometry is more rigid than that of the phenol. The thiazole analogue 6 was also designed based on tiazofurin that is converted to tiazofurin adenine dinucleotide (TAD) in vivo mimicking the NAD⁺ inverse regulation of IMPD. MPA side chain of this heterocyclic analogue may occupy the ribose phosphate portion of NAD⁺ in enzyme.

HO HO NH2 HO NH2 HO NH2 HO NH2
$$R = NH2$$
 $R = OMe$

According to molecular modeling studies in this laboratory, the superimposition of mycophenolic acid (MPA) and NAD⁺ shows that the MPA side chain, (E)-4-methyl-4-hexenoic acid, acts as a ribose phosphate equivalent. The carboxylate moiety of the MPA side chain is overlapped on the NAD⁺ phosphate, with the 2',3'-double bond being well aligned with the C4'-O4' bond of ribose. A similar relationship can be found in the structures of the naturally occurring neplanocins and the acyclic neplanocin analogues. Also, substitution of the phosphate group with isosteric carboxylic acids in drug design has been successful in other systems. Therefore, the ribotides of 6-chloropurine ribotides and ribavirin 5'-phosphate, IMP analogues as IMPD inhibitors, are replaced by the MPA side chain, (E)-4-methyl-4-hexenoic acid, to provide target compounds 7-10. These target compounds may bind to the IMP site in IMPD, and not to the NAD⁺ site, where MPA and tiazofurin bind.

As synthons of the MPA side chain, two allylic chlorides of (E)-4-methyl-4-hexenoic acid, 12 and 13, were synthesized from aldehyde 11 which was prepared from geraniol acetate by literature procedures. (Scheme 1). Synthesis of pyrrole analogues, 3 and 4, was accomplished via the direct incorporation of the MPA side chain synthon 12 on the pyrrole ring as a key step (Scheme 2). Alkylation of pyrrylmagnesium

Scheme 1

Reagent: (a) AgNO₃, KOH, EtOH-H₂O (b) CH₂N₂, ether (c) LiCl, MsCl, 2,6-lutidine, DMF, 0°C, (d) NaBH₄, EtOH (e) TBDMSCl, NEt₃, DMAP, CH₂Cl₂ (f) K₂CO₃, MeOH

Scheme 2

Reagent: (a) 12, toluene, -40 °C - r.t (b) KOH, EtOH-H₂O (c) Cl₃COCl, ether, 0 °C - r.t (d) NH₃, MeOH (e) NaOMe, MeOH

Scheme 3

13
$$\frac{\mathbf{a}, \mathbf{b}}{59 \%}$$
 TBDMSO NH₂ $\frac{\mathbf{c}}{56 \%}$ HO OE: $\frac{\mathbf{d}, \mathbf{e}}{71 \%}$ 6

Reagent: (a) LiCN, DMF, r.t (b) H_2S , KOH, MeOH, 90 °C (c) BrCH $_2$ C(=O)CO $_2$ Et, CH $_3$ CN, 0 °C - r.t (d) CrO $_3$, H_2SO_4 , acetone (e) NH $_3$, MeOH

Scheme 4

 $\textit{Reagent:} (a) \ 6\text{-chloropurine,} \ K_2CO_3, \ DMSO, \ r.t. \ (b) \ 0.5 \ N \ KOH, \ EtOH-H_2O, \ r.t \ (c) \ 0.1 \ N \ HCl, \ reflux$

Scheme 5

Reagent: (a) NaH, DMF, then 13 (b) Bu₄NF, THF (c) CrO₃, H₂SO₄, acetone (d) NH₃, MeOH (e) KOH, EtOH-H₂O

bromide 14¹² with allylic chloride 12 followed by ester hydrolysis, gave 2-alkylated pyrrole 15 as the major product (40%) along with 3-alkylated pyrrole (15%). Ring acylation of 15 with trichloroacetyl chloride¹³ gave the expected 2,5-disubstituted pyrrole 16 as the only regioisomer. 14 Ammonolysis of 16 gave the amide analogue 3, while treatment of 16 with sodium methoxide afforded the ester analogue 4. Synthesis of pyridine analogue 5 was previously described in detail.¹⁵ Synthesis of tiazofurin analogue 6 was based on the cyclization of the thioamide of the MPA side chain 13 with an α-halo ketone (Scheme 3). Allylic chloride 13 was converted to the corresponding nitrile, which was treated with potassium hydroxide and methanol-saturated hydrogen sulfide in a sealed bomb to give thioamide 17 without any deprotection of the silv group. Thioamide 17 was cyclized to the thiazole carboxylate 18 with ethyl bromopyruvate16, with simultaneous cleavage of the silyl protecting group under the acidic conditions. Oxidation of alcohol 18 followed by ammonolysis produced target compound 6. Two purine analogues, 7 and 8, were synthesized as IMP analogues by N-9 alkylation of 6-chloropurine with a MPA side chain synthon 12 (Scheme 4). Mild basic hydrolysis of N-9 alkylated 19 afforded 6-chloropurine analogue 7, while acid hydrolysis of 19 gave the inosine analogue 8. Synthesis of ribavirin analogues, 9 and 10, was based on the N-alkylation of methyl 1,2,4-triazole-3(5)-carboxylate 2018 with a synthon of the MPA side chain 13 (Scheme 5). Alkylation of 20 with allylic chloride 13 at room temperature afforded the 1,3-isomer 21 as the major isomer (56%) along with the 1,5-isomer (12%). Both structures were assigned based on a comparison of chemical shifts in each isomer according to a previous report.¹⁹ Deprotection of 21 followed by Jone's oxidation gave acid 22. Ammonolysis of 22 produced amide analogue 9, while basic hydrolysis of 22 gave acid analogue 10. All synthetic compounds were fully characterized by spectroscopy and elemental analysis.20

The lead compound, mycophenolic acid 1, and all final compounds 3-10 were tested *in vitro* against a number of organ specific human solid tumor cell lines using a sulforhodamine colormetric assay.²¹ IC₅₀ values of MPA are given in the parentheses: L1210 murine lymphocytic leukemia (<0.2 μ M), A549 human NSCLC lung carcinoma (0.4 μ M), HT-29 human colon adenocarcinoma (0.9 μ M), MCF7 human breast adenocarcinoma (0.5 μ M), and PSN-1 human pancreatic carcinoma (0.7 μ M). However, none of all prepared compounds showed a significant antitumor activity (IC₅₀ = > 100 μ M). MPA exhibited low activity (only 20

percent protection at 0.4 μ M) against HIV infected T4 lymphocytes (CEM cell line) and was cytotoxic to the uninfected cell with an IC₅₀ = 0.9 μ M. Among tested compounds, only compound 3 and 4 showed a moderate activity to T4 lymphocytes with IC₅₀ = 0.73 mM and 0.36 mM, respectively.

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- 20. Compound 3: white solid: mp 129-130 0 C; 1 H NMR (acetone-D6) δ 10.7 (bs, 1 H), 10.0 (bs, 1 H), 7.00 (bs, 2 H), 6.77 (m, 1 H), 5.90 (m, 1 H), 5.45 (t, 1 H), 3.40 (d, 2 H), 2.40 (m, 4 H), 1.70 (s, 3 H); IR (KBr) 3465 (NH), 3320, 3209 (OH), 2977, 1712 (C=O), 1633 (C=O), 1591, 1492, 1453, 1407, 1266 cm⁻¹. Anal. Calcd for $C_{12}H_{16}N_2O_3$: C, 61.00; H, 6.83; N, 11.86. Found: C, 60.85; H, 6.88; N, 11.79.

Compound 4: white solid: mp 128 $^{\circ}$ C; 1 H NMR (CDCl₃) δ 9.6 (bs, 1 H), 8.8 (bs, 1 H), 6.83 (m, 1 H), 5.93 (m, 1 H), 5.43 (t, 1 H), 3.80 (s, 3 H), 3.37 (d, 2 H), 2.50 (m, 4 H), 1.70 (s, 3 H); IR (KBr) 3301 (NH, OH), 2984, 1680 (C=O), 1488, 1331, 1302, 1210 cm⁻¹. Anal. Calcd for $C_{13}H_{17}NO_4$: C, 62.13; H, 6.82; N, 5.57. Found: C, 62.12; H, 6.86; N, 5.59.

Compound 6: white solid: mp 144-145 0 C; 1 H NMR (DMSO-d_o) δ 8.07 (s, 1 H), 7.3-7.8 (bd, 2 H), 5.50 (t, 1 H), 3.70 (d, 2 H), 2.1-2.6 (m, 4 H), 1.70 (s, 3 H); IR (KBr) 3365 (NH), 3107 (OH), 2921, 1702 (C=O), 1677 (C=O), 1658, 1521, 1475, 1304, 1213 cm⁻¹. Anal. Calcd for C₁₁H₁₄N₂O₃S: C, 51.95; H, 5.55; N, 11.01; S, 12.61. Found: C, 52.04; H, 5.58; N, 11.04; S, 12.58. Compound 7: white solid: mp 139-140 0 C; UV (EtOH): λ max 264.7 (ϵ 12,000); 1 H NMR (DMSO-d6) δ 8.80 (s, 1 H), 8.67 (s, 1 H), 5.52 (t, 1 H), 4.93 (d, 2 H), 2.1-2.6 (m, 4 H), 1.87 (s, 3 H).; IR (KBr) 3112 (OH), 3078, 2907, 2725, 2542, 1710 (C=O), 1599, 1570, 1451, 1404, 1347, 1244, 1176 cm⁻¹. Anal. Calcd for C₁₂H₁₃CIN₄O₂: C, 51.34; H, 4.67; Cl, 12.63; N, 19.96. Found: C, 51.44; H, 4.68; Cl, 12.59; N, 19.92.

Compound 8: white solid: mp decomp. >213 $^{\circ}$ C; UV (50% EtOH): λ max 249.9 (ϵ 12,800); 1 H NMR (DMSO-d6) δ 12-12.5 (bs, 1 H), 8.07 (bs, 2 H), 5.45 (t, 1 H), 4.70 (d, 2 H), 2.9-4.0 (m, 4 H), 1.82 (s, 3 H).; IR (KBr) 3121 (OH), 3057, 2914, 1713 (C=O), 1694, 1589, 1552, 1412, 1341, 1298, 1217, 1191, 1118 cm⁻¹. Anal. Calcd for $C_{12}H_{14}N_4O_3(+H_2O)$: C, 51.42; H, 5.76; N, 19.99. Found: C, 51.30; H, 5.78; N, 19.87.

Compound 9: white solid: mp 122-123 $^{\circ}$ C; 1 H NMR (DMSO-d6) δ 8.53 (s, 1 H), 7.53-7.8 (bd, 2 H), 5.43 (t, 1 H), 4.85 (d, 2 H), 2.2-2.6 (m, 4 H), 1.77 (s, 3 H).; IR (KBr) 3345 (NH), 3128 (OH), 3090, 1705 (C=O), 1660 (C=O), 1595, 1490, 1433, 1270 cm⁻¹. Anal. Calcd for $C_{10}H_{14}N_4O_3$: C, 50.41; H, 5.92; N, 23.52. Found: C, 50.36; H, 5.95; N, 23.34.

Compound 10: white solid: mp 169-170 0 C; 1 H NMR (DMSO-d6) δ 8.60 (s, 1 H), 7.35-7.8 (bd, 2 H), 5.43 (t, 1 H), 4.85 (d, 2 H), 2.1-2.6 (m, 4 H), 1.77 (s, 3 H).; IR (KBr) 3113 (OH), 1697 (C=O) 1653 (C=O), 1648, 1446 cm⁻¹. Anal. Calcd for $C_{10}H_{13}N_{3}O_{4}$: C, 50.20; H, 5.48; N, 17.57. Found: C, 50.14; H, 5.51; N, 17.54.

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